

## HUMAN TOTAL CATHEPSIN S ELISA KIT

FOR THE QUANTITATIVE DETERMINATION OF  
HUMAN TOTAL CATHEPSIN S  
CONCENTRATIONS IN CELL CULTURE  
SUPERNATES, SERUM, AND PLASMA



FOR RESEARCH USE ONLY. NOT FOR USE  
IN DIAGNOSTIC PROCEDURES.

### PURCHASE INFORMATION:

ELISA NAME	HUMAN TOTAL CATHEPSIN S ELISA
Catalog No.	SK00371-01
Lot No.	
Formulation	96 T
Standard Range	15.6 - 1000 pg/mL
Sensitivity	4 pg/mL
Sample Volume	100 µL
Dilution Factor	20 (Optimal dilutions should be determined by each laboratory for each application)
Sample Type	Serum, EDTA Plasma, Cell Culture Supernates
Specificity	Human Total Cathepsin S including pro, mature, and cystatin-complexed forms
Intra-assay Precision	4 - 8%
Inter-assay Precision	8 - 10%
Storage	2 – 8 °C

### Order Contact:

AVISCERA BIOSCIENCE, INC.  
2348 WALSH AVE., SUITE C  
SANTA CLARA, CA 95051  
USA

Tel: 408-982-0300

Fax: 408-982-0301

Email: [Info@AvisceraBioscience.com](mailto:Info@AvisceraBioscience.com)

[Sales@AvisceraBioscience.com](mailto:Sales@AvisceraBioscience.com)

[www.AvisceraBioscience.com](http://www.AvisceraBioscience.com)

## INTRODUCTION

Human Cathepsin S immunoassay is a solid phase ELISA designed to measure human Cathepsin S in cell culture supernates, serum, and plasma. It contains recombinant human Cathepsin S and antibodies raised against this protein. It has been shown to accurately quantify recombinant human Cathepsin S. Results obtained with naturally occurring Cathepsin S samples showed linear curves that were parallel to the standard curves obtained using the kit standards. These results indicate that the immunoassay kit can be used to determine relative mass values for natural human Cathepsin S.

## PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. An antibody specific for Cathepsin S has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any Cathepsin S present is bound by the immobilized antibody. After washing away any unbound substances, a biotinylated antibody specific for Cathepsin S is added to the wells. Following a wash to remove any unbound antibody-biotin reagent, HRP link Streptavidin is added to the wells. After washing away any unbound enzyme, a substrate solution is added to the wells and color develops in proportion to the amount of Cathepsin S bound in the initial step. The color development is stopped and the intensity of the color is measured.

## LIMITATIONS OF THE PROCEDURE

\_ FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.

\_ The kit should not be used beyond the expiration date on the kit label.

\_ Do not mix or substitute reagents with those from other lots or sources.

\_ It is important that the Dilution Buffer selected for the standard curve be consistent with the samples being assayed.

\_ If samples generate values higher than the highest standard, dilute the samples with Dilution Buffer and repeat the assay.

\_ Any variation in standard diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.

\_ This assay is designed to eliminate interference by soluble receptors, binding proteins, and other factors present in biological samples. Until all factors

have been tested in the immunoassay, the possibility of interference cannot be excluded.

## MATERIALS PROVIDED

DESCRIPTION	CODE	QUANTITY
<b>Cathepsin S Microplate</b> - 96 well polystyrene microplate (12 strips of 8 wells) coated with an antibody against Cathepsin S.	<b>371-01-01</b>	<b>1 plate</b>
<b>CATHEPSIN S Standard</b> – 1000 pg/vial of recombinant human Cathepsin S in a buffered protein base with preservative; lyophilized.	<b>371-01-02</b>	<b>1 vial</b>
<b>Detection Antibody Concentrate</b> – 1.05 mL/vial, 10-fold concentrate of biotinylated antibody against Cathepsin S with preservative; lyophilized.	<b>371-01-03</b>	<b>1 vial</b>
<b>Positive Control</b> – one vial of recombinant Cathepsin S; lyophilized.	<b>371-01-04</b>	<b>1 vial</b>
<b>Streptavidin-HRP Conjugate</b> - 60 µL/vial, 200-fold concentrated solution of Streptavidin conjugate to HRP with preservative.	<b>SAHRP</b>	<b>1 vial</b>
<b>Dilution Buffer</b> – 60 mL of buffered protein based solution with preservative.	<b>DB01</b>	<b>1 bottle</b>
<b>Antibody Diluent Solution</b> – 12 mL of buffered protein based solution with preservative.	<b>DB08</b>	<b>1 bottle</b>
<b>Wash Buffer</b> - 50 mL of 10-fold concentrated buffered surfactant, with preservative.	<b>WB01</b>	<b>1 bottle</b>
<b>TMB Substrate Solution</b> - 11 mL of TMB substrate solution.	<b>TMB01</b>	<b>1 bottle</b>
<b>Stop Solution</b> – 11 mL of 0.5M HCl.	<b>S-STOP</b>	<b>1 bottle</b>
<b>Plate Sealer</b>	<b>EAPS</b>	<b>1 piece</b>
<b>Plastic Pouch</b>	<b>P01</b>	<b>1 piece</b>

## STORAGE

**Unopened Kit:** Store at 2 – 8 °C for up to 12 months. For longer storage, unopened Standard, Positive Control and Detection Antibody Concentrate should

be stored at -20 °C or -70 °C. Do not use kit past expiration date.

**Opened / Reconstituted Reagents:** Reconstituted Standard (stock) solution and Detection Antibody concentrated solution SHOULD BE STORED at -20 °C or -70 °C for up to one month.

Streptavidin-HRP Conjugate 200-fold concentrated solution (protect from light) and other components may be stored at 2 – 8 °C for up to 12 months.

**Microplate Wells:** Return unused wells to the plastic pouch with the desiccant pack. Microplate may be stored for up to 6 months at 2 – 8 °C after opening.

### OTHER SUPPLIES REQUIRED

- Microplate reader capable of measuring absorbance at 450 nm.
- Microplate shaker (250-300rpm).
- Pipettes and pipette tips.
- Deionized or distilled water.
- Squirt bottle, manifold dispenser, or automated microplate washer.
- 100 mL and 500 mL graduated cylinders.

### SAMPLE COLLECTION AND STORAGE

**Cell Culture Supernates** - Remove particulates by centrifugation and assay immediately or aliquot and store samples at  $\leq -20^{\circ}\text{C}$ . Avoid repeated freeze-thaw cycles.

**Serum** - Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at  $\leq -20^{\circ}\text{C}$ . Avoid repeated freeze-thaw cycles.

**Plasma** - Collect plasma using EDTA, heparin, or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at  $\leq -20^{\circ}\text{C}$ . Avoid repeated freeze-thaw cycles.

**Optional:** Use Aprotinin (enzyme inhibitor) for ALL sample collection to prevent sample degradation. 0.5 TIU per ml of sample solution.

### SAMPLE PREPARATION

Serum and plasma samples may require a 20-fold dilution. A suggested 20-fold dilution is 12  $\mu\text{L}$  sample + 228  $\mu\text{L}$  Dilution Buffer.

**Optimal dilutions should be determined by each laboratory for each application with a pretest.**

Use polypropylene test tubes.

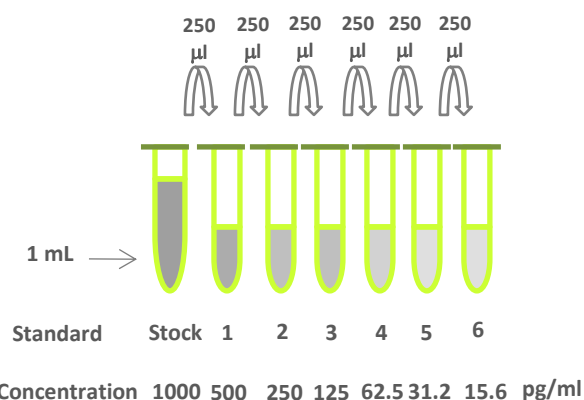
### REAGENT PREPARATION

**Bring all reagents to room temperature before use.**

**Wash Buffer** - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 50 mL of Wash Buffer Concentrate into deionized or distilled water (450 mL) to prepare 500 mL of 1x Wash Buffer.

**Cathepsin S Standard - Refer to vial label for reconstitution volume.** Reconstitute the **Cathepsin S** standard with 1.0 mL of Dilution Buffer. This reconstitution produces a stock solution of 1000 pg/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Pipette 250  $\mu\text{L}$  of Dilution Buffer into tubes #1 to #6. Use the stock solution to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The 1000 pg/mL standard serves as the high standard. The Dilution Buffer serves as the zero standard (0 pg/mL).

Tube	Standard	Dilution Buffer	Concentration
stock	powder	1.0 ml	1000 pg/ml
# 1	250 $\mu\text{L}$ of stock	250 $\mu\text{L}$	500 pg/ml
# 2	250 $\mu\text{L}$ of 1	250 $\mu\text{L}$	250 pg/ml
# 3	250 $\mu\text{L}$ of 2	250 $\mu\text{L}$	125 pg/ml
# 4	250 $\mu\text{L}$ of 3	250 $\mu\text{L}$	62.5 pg/ml
# 5	250 $\mu\text{L}$ of 4	250 $\mu\text{L}$	31.25 pg/ml
# 6	250 $\mu\text{L}$ of 5	250 $\mu\text{L}$	15.6 pg/ml



**Positive Control** - Reconstitute the positive control with 1.0 mL of Dilution Buffer to make positive control working solution. **Note:** Positive control could be reused within a few days if stored at -20 °C or -70 °C.

**Detection Antibody Concentrate** - Reconstitute the Detection Antibody Concentrate with 1.05 mL of **Antibody Diluent Solution (DB08)** to produce a 10-fold concentrated stock solution. Pipette 9.45 mL of Antibody Diluent Solution (DB08) into a 15 mL centrifuge tube and transfer 1.05 mL of 10-fold concentrated stock solution to prepare working solution.

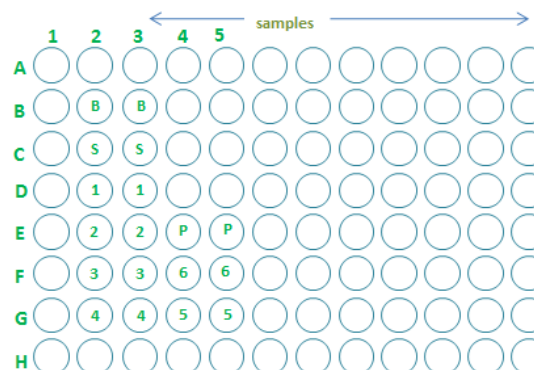
**Streptavidin-HRP Conjugate** - Pipette 11.94 mL of Dilution Buffer into a 15 mL centrifuge tube and transfer 60  $\mu$ L of 200-fold concentrated stock solution to prepare working solution. **Note:** 1x working solution of Streptavidin-HRP Conjugate should be used within a few days (**protect from light**).

## ASSAY PROCEDURE

Bring all reagents and samples to room temperature before use. It is recommended that blank, standards, positive control and samples be assayed in duplicate.

1. Prepare all reagents and working standards as directed in the previous sections.
2. Remove excess microplate strips from the plate frame, return them to the plastic pouch with the desiccant pack.
3. Add 100  $\mu$ L of Dilution Buffer to Blank wells (B2, B3).
4. Add 100  $\mu$ L of Standard solutions in reverse order of serial dilution (F4, F5 to G4, G5 and G2, G3 to C2, C3), sample, or positive control (E4, E5) per well. Cover with plate sealer. Incubate for 2 hours on microplate shaker at room temperature. A plate layout is provided to record standards and samples assayed.
5. Aspirate each well and wash, repeating the process three times for a total of four washes. Wash by filling each well with 1x Wash Buffer (300  $\mu$ L) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
6. Add 100  $\mu$ L of Detection Antibody working solution to each well. Cover with plate sealer. Incubate for 2 hours on microplate shaker at room temperature.
7. Repeat the aspiration/wash as in step 5.

8. Add 100  $\mu$ L of Streptavidin-HRP Conjugate working solution to each well. Incubate for 45 minutes on microplate shaker at room temperature. **Protect from light.**
9. Repeat the aspiration/wash as in step 5.
10. Add 100  $\mu$ L of Substrate Solution to each well. Incubate for 3-6 minutes microplate shaker at room temperature. **Protect from light.**
11. Add 100  $\mu$ L of Stop Solution to each well. The color in the wells should change from blue to yellow. If the color in the wells is green, or if the color change does not appear uniform, gently tap the plate to ensure thorough mixing.
12. Determine the optical density of each well within 15 minutes, using a microplate reader set to 450 nm.



## CALCULATION OF RESULTS

Average the duplicate readings for each standard, positive control and sample, and subtract the average zero standard optical density. Create a standard curve by reducing the data using computer software capable of generating a log-log curve fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the Cathepsin S concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.

If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

## CALIBRATION

This immunoassay is calibrated against a highly purified recombinant human Cathepsin S.

**TYPICAL DATA**

This standard curve data is provided for demonstration only. A standard curve should be generated for each set of samples assayed.

CATHEPSIN S (PG/ML)	CORRECTED (450NM)
Blank	0 (0.103)
7.813 (optional)	0.043
15.625	0.083
31.25	0.155
62.5	0.289
125	0.559
250	1.053
500	1.881
1000	2.872

- Lot No.:
- Positive Control :

**SENSITIVITY**

The minimum detectable dose (MDD) of Cathepsin S was 4 pg/mL.

**SPECIFICITY**

This assay recognizes both natural and recombinant human Cathepsin S. The factors listed below were prepared at 50 ng/mL in Dilution Buffer, and assayed for cross reactivity. Preparations of the following factors at 50 ng/mL in a mid-range rh Cathepsin S control were assayed for interference. No significant cross-reactivity or interference was observed.

	Cross-Reactivity (%)
Human Cathepsin S	100
Human Cathepsin A	0
Human Cathepsin B	0
Human Cathepsin D	0
Human Cathepsin E	0
Human Cathepsin L	0
Human Cathepsin V	0

This kit detects total Cathepsin S including the pro, mature, and cystatin-complexed forms.

**SUMMARY OF ASSAY PROCEDURE****PREPARE REAGENTS, SAMPLES AND STANDARDS**

↓  
Add 100 µl of standard, samples, positive control to each well. Incubate 2 hours on the plate shaker at RT.

↓  
Aspirate and wash 4 times.

↓  
Add 100 µl Detection Antibody working solution to each well. Incubate 2 hours on the plate shaker at RT.

↓  
Aspirate and wash 4 times.

↓  
Add 100 µl Streptavidin-HRP conjugate working solution to each well. Incubate 45 minutes on the plate shaker at RT. **Protect from light.**

↓  
Aspirate and wash 4 times.

↓  
Add 100 µl Substrate Solution to each well. Incubate 3-6 minutes on the plate shaker at RT. **Protect from light.**

↓  
Add 100 µl Stop Solution to each well. Read 450nm within 15 minutes.