

HUMAN sP-SELECTIN ELISA KIT

FOR THE QUANTITATIVE DETERMINATION OF HUMAN sP-SELECTIN CONCENTRATIONS IN CELL CULTURE SUPERNATES, SERUM AND PLASMA



FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.

PURCHASE INFORMATION:

ELISA NAME	HUMAN sP-SELECTIN ELISA
Catalog No.	SK00093-02
Lot No.	
Formulation	96 T
Standard range	93.75-6000 pg/mL
Sensitivity	47 pg/mL
Sample Volume	100 µL
Dilution Factor	40x (Optimal dilutions should be determined by each laboratory for each application)
Sample Type	Plasma, Serum, Cell Culture Supernates
Specificity	Human sP-Selectin
Intra-assay Precision	4-6%
Inter-assay Precision	8-10%
Storage	2°C - 8°C

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INTRODUCTION

Human sP-SELECTIN immunoassay is a 3.5 - 4.5 hour solid phase ELISA designed to measure human sP-SELECTIN in cell culture supernates, serum, and plasma. It contains recombinant human sP-SELECTIN and antibodies raised against this protein. It has been shown to accurately quantify recombinant human sP-SELECTIN. Results obtained with naturally occurring sP-SELECTIN samples showed linear curves that were parallel to the standard curves obtained using the kit standards. These results indicate that the immunoassay kit can be used to determine relative mass values for natural human sP-SELECTIN.

PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. A monoclonal antibody specific for sP-SELECTIN has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any sP-SELECTIN present is bound by the immobilized antibody. After washing away any unbound substances, a biotinylated polyclonal antibody specific for sP-SELECTIN is added to the wells. Following a wash to remove any unbound antibody-biotin reagent, HRP link Streptavidin is added to the wells. After washing away any unbound enzyme, a substrate solution is added to the wells and color develops in proportion to the amount of sP-SELECTIN bound in the initial step. The color development is stopped and the intensity of the color is measured.

LIMITATIONS OF THE PROCEDURE

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_ The kit should not be used beyond the expiration date on the kit label.

_ Do not mix or substitute reagents with those from other lots or sources.

_ It is important that the Dilution Buffer selected for the standard curve be consistent with the samples being assayed.

_ If samples generate values higher than the highest standard, dilute the samples with Dilution Buffer and repeat the assay.

_ Any variation in standard diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.

_ This assay is designed to eliminate interference by soluble receptors, binding proteins, and other factors present in biological samples. Until all factors

have been tested in the immunoassay, the possibility of interference cannot be excluded.

MATERIALS PROVIDED

DESCRIPTION	CODE	QUANTITY
sP-SELECTIN Microplate - 96 well polystyrene microplate (12 strips of 8 wells) coated with a monoclonal purified IgG against human sP-SELECTIN.	093-02-01	1 plate
sP-SELECTIN Standard – 6000 pg/vial of recombinant human sP-SELECTIN in a buffered protein base with preservatives; lyophilized.	093-02-02	1 vial
Detection Antibody Concentrate – 105 µL/vial, 100-fold concentrated of biotinylated polyclonal purified IgG against human sP-SELECTIN with preservatives; lyophilized.	093-02-03	1 vial
Positive Control – one vial of recombinant human sP-SELECTIN, lyophilized	093-02-04	1 vial
Streptavidin-HRP Conjugate – 60 µL/vial, 200-fold concentrated solution of Streptavidin conjugate to HRP with preservatives	SAHRP	1 vial
Dilution Buffer - 60mL of buffered protein based solution with preservatives	DB01	1 bottle
Wash Buffer - 50 mL of 10-fold concentrated buffered surfactant, with preservative.	WB01	1 bottle
TMB Substrate Solution - 11 mL of TMB substrate solution	TMB01	1 bottle
Stop Solution - 11 mL of 0.5M HCl	S-STOP	1 bottle
Plate Sealer	EAPS	1 piece

STORAGE

Unopened Kit: Store at 2 - 8°C for up to 6 months. For longer storage, unopened Standard, Positive Control and Detection Antibody Concentrate should be stored at -20°C or -70°C. Do not use kit past expiration date.

Opened / Reconstituted Reagents: Reconstituted Standard and Antibody Solution SHOULD BE STORED at -20°C or -70°C for up to one month. Streptavidin-HRP Conjugate 200-fold concentrate and other components may be stored at 2 - 8°C for up to 6 months.

Microplate Wells: Return unused wells to the plastic pouch containing the desiccant pack, reseal along entire edge of zip-seal. Microplate may be stored for up to 6 months at 2 - 8°C.

OTHER SUPPLIES REQUIRED

- Microplate reader capable of measuring absorbance at 450 nm.
- Microplate shaker (250-300rpm).
- Pipettes and pipette tips.
- Deionized or distilled water.
- Squirt bottle, manifold dispenser, or automated microplate washer.
- 100 mL and 500 mL graduated cylinders.

SAMPLE COLLECTION AND STORAGE

Cell Culture Supernates - Remove particulates by centrifugation and assay immediately or aliquot and store samples at ≤-20°C or -70°C. Avoid repeated freeze-thaw cycles.

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at ≤ -20°C or -70°C. Avoid repeated freeze-thaw cycles.

Plasma - Collect plasma using EDTA, heparin, or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at ≤-20°C or -70°C. Avoid repeated freeze-thaw cycles.

Note: Use Aprotinin (enzyme inhibitor) (Code No.: 00700-01-25) for ALL sample collection to prevent sample degradation. 0.5 TIU per ml of sample solution.

SAMPLE PREPARATION

Serum and plasma samples may require a 40-fold dilution. A suggested 40-fold dilution is 10 µL sample + 390 µL Dilution Buffer.

Optimal dilutions should be determined by each laboratory for each application.

Use polypropylene test tubes.

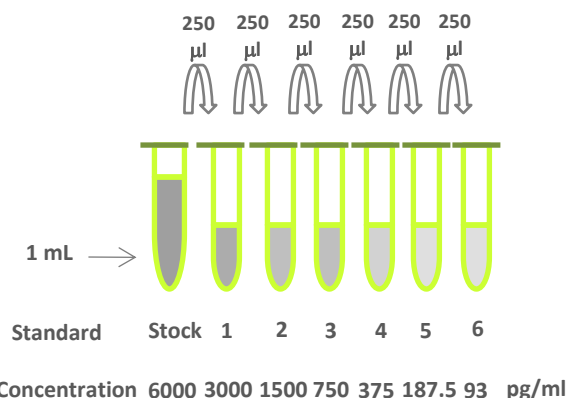
REAGENT PREPARATION

Bring all reagents to room temperature before use.

Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 50 mL of Wash Buffer Concentrate into deionized or distilled water (450 mL) to prepare 500 mL of Wash Buffer.

sP-SELECTIN Standard - Refer to vial label for reconstitution volume. Reconstitute the sP-SELECTIN standard with 1.0 mL of Dilution Buffer. This reconstitution produces a stock solution of 6000 pg/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Pipette 250 µL of Dilution Buffer into tubes #1 to #6. Use the stock solution to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The 6000 pg/mL standard serves as the high standard. The Dilution Buffer serves as the zero standard (0 pg/mL).

TUBE	STANDARD	DILUTION BUFFER	CONCENTRATION
stock	powder	1.0 mL	6000 pg/ml
# 1	250µl of stock	250µl	3000 pg/ml
# 2	250µl of 1	250µl	1500 pg/ml
# 3	250µl of 2	250µl	750 pg/ml
# 4	250µl of 3	250µl	375 pg/ml
# 5	250µl of 4	250µl	187.5 pg/ml
# 6	250µl of 5	250µl	93.75 pg/ml



Detection Antibody Concentrate - Reconstitute the Detection Antibody Concentrate with 105 μ L of Dilution Buffer to produce a 100-fold concentrated stock solution. Pipette 10.395 mL of Dilution Buffer into a 15 mL centrifuge tube and transfer 105 μ L of 100-fold concentrated stock solution to prepare working solution.

Positive Control - Reconstitute the positive control with 1.0 mL of Dilution Buffer to make positive control solution. **Note:** Positive control should be used within a few days.

Streptavidin-HRP Conjugate - Pipette 11.94 mL of Dilution Buffer into a 15 mL centrifuge tube and transfer 60 μ L of 200-fold concentrated stock solution to prepare working solution. **Note:** 1x working solution of Streptavidin-HRP Conjugate should be used within a few days.

ASSAY PROCEDURE

Bring all reagents and samples to room temperature before use. It is recommended that blank, standards and positive control be assayed in duplicates.

1. Prepare all reagents and working standards as directed in the previous sections.
2. Remove excess micro-plate strips from the plate frame, return them to the plastic pouch containing the desiccant pack, reseal.
3. Add 100 μ L of **Dilution Buffer** to Blank wells (F4, F5).
4. Add 100 μ L of **Standard** (B2, B3 to G2, G3 and G4, G5), **sample**, or **positive control** (E4, E5) per well. Cover with plate sealer. Incubate for 2 hours on micro-plate shaker at room temperature. A plate layout is provided to record standards and samples assayed.
5. Aspirate each well and wash, repeating the process three times for a total of four washes. Wash by filling each well with **1x Wash Buffer** (300 μ L) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
6. Add 100 μ L of **Detection Antibody working solution** to each well. Cover with sealer. Incubate

for 2 hours on micro-plate shaker at room temperature.

7. Repeat the aspiration/wash as in step 5.
8. Add 100 μ L of **Streptavidin-HRP Conjugate working solution** to each well. Incubate for 1 hour on micro-plate shaker at room temperature.
Protect from light.
9. Repeat the aspiration/wash as in step 5.
10. Add 100 μ L of **Substrate Solution** to each well. Incubate for 2-5 minutes at room temperature.
Protect from light.
11. Add 100 μ L of **Stop Solution** to each well. The color in the wells should change from blue to yellow. If the color in the wells is green, or if the color change does not appear uniform, gently tap the plate to ensure thorough mixing.
12. Determine the optical density of each well within 15 minutes, using a micro-plate reader set to 450 nm.

CALCULATION OF RESULTS

Average the duplicate readings for each standard, positive control, and sample and subtract the average zero standard optical density. Create a standard curve by reducing the data using computer software capable of generating a log-log curve fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the sP-SELECTIN concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.

If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

CALIBRATION

This immunoassay is calibrated against a highly purified CHO-expressed recombinant human sP-SELECTIN.

SPECIFICITY

The factors listed below were prepared at 50 ng/mL in Dilution Buffer, and assayed for cross reactivity. No significant cross-reactivity or interference was observed.

PROTEINS	CROSS-REACTIVITY (%)
Human P-Selectin	100
Mouse P-Selectin	0.4
Human E-Selectin	0
Human L-Selectin	0

TYPICAL DATA

The standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.

sP-SELECTIN (PG/ML)	AVERAGE OD450 (CORRECTED)
Blank	0 (0.056)
46.8 (optional)	0.021
93.75	0.049
187.5	0.105
375	0.227
750	0.425
1500	0.652
3000	1.047
6000	1.452

- Lot No.:
- Positive Control: 400 – 1000 pg/mL

SUMMARY OF ASSAY PROCEDURE**PREPARE REAGENTS, SAMPLES AND STANDARDS**

Add 100 µL of **standard, samples, positive control** to each well. Incubate 2 hours on plate shaker at RT.

Aspirate and wash 4 times.

Add 100 µL **Detection Antibody working solution** to each well. Incubate 2 hours on plate shaker at RT.

Aspirate and wash 4 times.

Add 100 µL **Streptavidin-HRP conjugate working solution** to each well. Incubate 1 hour on plate shaker at RT. **Protect from light.**

Aspirate and wash 4 times.

Add 100 µL **Substrate Solution** to each well. Incubate 2-5 min on plate shaker. **Protect from light.**

Add 100 µL **Stop Solution** to each well. Read 450nm within 15 min