

HIGH SENSITIVITY TISSUE INHIBITOR OF METALLOPROTEINASES 1 (TIMP-1) (HUMAN) ELISA KIT

FOR THE QUANTITATIVE DETERMINATION
OF HUMAN TIMP-1 CONCENTRATIONS IN
SERUM AND EDTA PLASMA



ALWAYS REFER TO LOT SPECIFIC PROTOCOL
PROVIDED WITH EACH KIT FOR
INSTRUCTIONS. PROTOCOL MUST BE
READ BEFORE USING THIS PRODUCT.

FOR RESEARCH USE ONLY. NOT FOR USE IN
DIAGNOSTIC PROCEDURES.

PRODUCT INFORMATION:

THIS KIT IS FOR ONE TIME USE ONLY.

ELISA NAME	HIGH SENSITIVITY TIMP-1 (HUMAN) ELISA
Catalog No.	SK00039-06
Lot No.	
Formulation	96 T
Standard range	62.5-1000 pg/mL
Sensitivity	15 pg/mL
Sample Volume	100 µl
Sample Type	Serum, EDTA Plasma
Dilution Factor	100 to 200 (Optimal dilutions should be determined by each laboratory for each application)
Specificity	Human TIMP-1
Intra-assay Precision	4-6%
Inter-assay Precision	8-12%
Storage	2°C - 8°C for 1 month. See page 2 for more information.
This kit contains sufficient materials to run approximately 35 samples duplicated provided that assay is run according to protocol.	

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INTRODUCTION

High Sensitivity TIMP-1 (Human) ELISA Kit contains the necessary components required for the quantitative measurement of recombinant and/or natural human TIMP-1 from serum or Plasma in a sandwich ELISA format.

This immunoassay contains recombinant human TIMP-1 and antibodies raised against this protein. Results from this immunoassay have shown to accurately quantify recombinant and natural TIMP-1 samples

PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. An antibody specific for human TIMP-1 has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any human TIMP-1 present is bound by the immobilized antibody. After washing away any unbound substances, a HRP conjugated antibody specific for human TIMP-1 is added to the wells. Following a wash to remove any unbound antibody- HRP conjugate, a substrate solution is added to the wells and color develops in proportion to the amount of TIMP-1 bound in the initial step. The color development is stopped and the intensity of the color is measured.

LIMITATIONS OF THE PROCEDURE

_FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.

_The kit should not be used beyond the expiration date on the kit label.

_Do not mix or substitute reagents with those from other lots or sources.

_It is important that the Dilution Buffer selected for the standard curve be consistent with the samples being assayed.

_If samples generate values higher than the highest standard, dilute the samples with Dilution Buffer and repeat the assay.

_ Any variation in standard diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.

_This assay is designed to eliminate interference by soluble receptors, binding proteins, and other factors present in biological samples. Until all factors have been tested in the immunoassay, the possibility of interference cannot be excluded.

MATERIALS PROVIDED

DESCRIPTION	CODE	QUANTITY
TIMP-1 Microplate - 96 well polystyrene microplate (12 strips of 8 wells) coated with an antibody against TIMP-1.	039-01-01	1 plate
TIMP-1 Standard – 2000 pg/vial of recombinant human TIMP-1 in a buffered protein base with preservatives; lyophilized.	039-06-02	1 vial
Detection Antibody HRP – 30 µL /vial, 400-fold concentrate of antibody HRP conjugate against TIMP-1 with preservatives; lyophilized.	039-01-03	1 vial
Dilution Buffer - 40mL of buffered protein based solution with preservatives	DB06	1 bottle
Antibody Diluent Solution – 12 mL of buffered protein based solution with preservative.	DB08B	1 bottle
Wash Buffer - 50mL of 10-fold concentrated buffered surfactant, with preservative.	WB01	1 bottle
TMB Substrate Solution - 11mL of TMB substrate solution	TMB01	1 bottle
Stop Solution - 11mL of 0.5M HCl	S-STOP	1 bottle
Plate Sealer	EAPS	1
Plastic Pouch	P01	1

STORAGE

Unopened Kit: Store at 2 – 8° C for up to 1 month. For longer storage up to 10 months, unopened Standard, Positive Control, Detection Antibody Concentrate, Dilution Buffer and HRP Diluent Solution should be stored at -20° C. Detection Antibody-HRP Conjugate and TMB Substrate Solution should be stored only at 2-8° C. Do not use kit past expiration date.

OTHER SUPPLIES REQUIRED

- Microplate reader capable of measuring absorbance at 450 nm.
- Microplate shaker (250-300rpm).
- Pipettes and pipette tips.
- Deionized or distilled water.

- Squirt bottle, manifold dispenser, or automated microplate washer.
- 100 mL and 500 mL graduated cylinders.

SAMPLE COLLECTION AND STORAGE

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

Plasma - Collect plasma using EDTA, heparin, or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at $\leq -20^{\circ}\text{C}$. Avoid repeated freeze-thaw cycles.

Note: Use Aprotinin (enzyme inhibitor) for ALL sample collection to prevent sample degradation. 0.5 TIU per ml of sample solution.

SAMPLE PREPARATION

Serum or EDTA plasma samples may require a 100 to 200-fold dilution. A suggested 100-fold dilution is 10 μL sample + 90 μL Dilution Buffer, followed by 25 μL of 10-fold diluted sample + 225 μL Dilution Buffer. A suggested 200-fold dilution is 10 μL sample + 90 μL Dilution Buffer, followed by 12 μL of 10-fold diluted sample + 228 μL Dilution Buffer. **Optimal dilutions should be determined by each laboratory for each application.**

Use polypropylene test tubes.

REAGENT PREPARATION

Bring all reagents to room temperature before use.

Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 50 mL of Wash Buffer Concentrate into deionized or distilled water (450 mL) to prepare 500 mL of Wash Buffer.

Dilution Buffer (DB06) - Dilution Buffer (DB06) is highly viscous, prior to use warm it in 30 - 37 $^{\circ}$ C water bath until liquid flows more freely.

TIMP-1 Standard - Refer to vial label for reconstitution volume. Reconstitute the **TIMP-1 Standard** with 1 mL of Dilution Buffer. This reconstitution produces a stock solution of 2000 pg/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Pipette 250 μL of Dilution Buffer into tubes

#1 to #5. Use the stock solution to produce a dilution series (below). Mix each tube thoroughly before the next transfer. The **1000 pg/mL** standard serves as the high standard. The Dilution Buffer serves as the zero standard (0 pg/mL).

TUBE	STANDARD	DILUTION BUFFER	CONCENTRATION
stock	Powder	1000 μL	2000 pg/ml
# 1	250 μL of stock	250 μL	1000 pg/ml
# 2	250 μL of 1	250 μL	500 pg/ml
# 3	250 μL of 2	250 μL	250 pg/ml
# 4	250 μL of 3	250 μL	125 pg/ml
# 5	250 μL of 4	250 μL	62.5 pg/ml
optional	250 μL of 5	250 μL	31.25 pg/ml

Detection Antibody HRP - Pipette 10.473 mL of **Antibody Diluent Solution (DB08B)** into a 15 mL centrifuge tube and transfer 27 μL of 400-fold concentrated stock solution to prepare working solution. **Note:** 1X working solution of Detection Antibody-HRP Conjugate should be used within a few hours. **Protect from light.**

ASSAY PROCEDURE

Bring all reagents and samples to room temperature before use. It is recommended that blank, standards, positive control and be assayed in duplicate.

1. Prepare all reagents and working standards as directed in the previous sections.
2. Remove excess micro-plate strips from the plate frame, return them to the plastic pouch with the desiccant pack and seal.
3. Add 100 μL of **Dilution Buffer** to Blank wells (E4, E5).
4. Add 100 μL of **Standard** (from B2, B3 to G2, G3 and G4, G5), **sample, or positive control** (F4, F5) per well. Cover with plate sealer. Incubate for 2 hours on micro-plate shaker at room temperature. A plate layout is provided to record standards and samples assayed.
5. Aspirate each well and wash, repeating the process three times for a total of four washes. Wash by filling each well with **1X Wash Buffer** (300 μL) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by

aspirating or decanting. Invert the plate and blot it against clean paper towels.

6. Add 100 μ L of **Detection Antibody working solution** to each well. Cover with plate sealer. Incubate for 1 hour on micro-plate shaker at room temperature.
7. Repeat the aspiration/wash as in step 5.
8. Add 100 μ L of **Substrate Solution** to each well. Incubate for 10-15 minutes on a micro-plate shaker at room temperature. **Protect from light.**
9. Add 100 μ L of **Stop Solution** to each well. The color in the wells should change from blue to yellow. If the color in the wells is green, or if the color change does not appear uniform, gently tap the plate to ensure thorough mixing.
10. Determine the optical density of each well within 3 minutes, using a micro-plate reader set to 450 nm.

CALCULATION OF RESULTS

Average the duplicate readings for each standard, positive control and sample, and subtract the average zero standard optical density. Create a standard curve by reducing the data using computer software capable of generating a log-log curve fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the TIMP-1 concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.

If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

CALIBRATION

This immunoassay is calibrated against a highly purified recombinant human TIMP-1.

SENSITIVITY

Twenty-five assays were evaluated and the minimum detectable dose (MDD) of TIMP-1 was 15pg/mL.

TYPICAL DATA

This standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.

Standard (pg/mL)	Average OD450nm (Corrected)
Blank	0 (0.132)
31.25 (optional)	0.038
62.5	0.079
125	0.157
250	0.331
500	0.520
1000	1.023
2000 (optional)	1.859

LINEARITY

To assess the linearity of the assay, pooled research human **EDTA plasma** samples were diluted with Dilution Buffer and assayed.

DILUTION FACTOR	ASSAYED (PG/ML)	FINAL (PG/ML)	RECOVERY (%)
200X	438.038	87607.6	100
400X	225.394	90157.6	103

To assess the linearity of the assay, pooled research human **serum** samples were diluted with Dilution Buffer and assayed.

DILUTION FACTOR	ASSAYED (PG/ML)	FINAL (PG/ML)	RECOVERY (%)
200X	2121.595	424319	100
400X	1380.383	552153	130

SPECIFICITY

This assay recognizes both natural and recombinant human TIMP-1. The factors listed below were prepared at 50 ng/mL in Dilution Buffer, and assayed for cross reactivity. Preparations of the following factors at 50 ng/mL in a mid-range rh TIMP-1 control were assayed for interference. No significant cross-reactivity or interference was observed.







Human Recombinant Proteins:

MMP-1, MMP-2, MMP-3, TIMP-2

Mouse Recombinant Proteins:

TIMP-1

SUMMARY OF ASSAY PROCEDURE

PREPARE REAGENTS, SAMPLES AND STANDARDS

Add 100 µl of standard, samples, positive control to each well. Incubate 2 hours on the plate shaker at RT.

Aspirate and wash 4 times.

Add 100 µl Detection Antibody working solution to each well. Incubate 1 hour on the plate shaker at RT.

Aspirate and wash 4 times.

Add 100 µl Substrate Solution to each well. Incubate 10-15 min on the plate shaker. Protect from light.

Add 100 µl Stop Solution to each well. Read 450nm within 3min